

**ALL INDIA COORDINATED RICE IMPROVEMENT
PROJECT
(AICRIP)**

**Entomology Technical Programme
*Kharif 2019 & Rabi 2019-20***



**ICAR - Indian Institute of Rice Research
Rajendranagar, Hyderabad 500 030**

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Trial Star Sheet - Kharif 2019

Sl.No.	Location	PSR\$	PHS	GMS	GMSS	LFST	SBST	MRST	NSN-1	NSN-2	NSN-H	NHSN	GMBT	PHSS	GMPM	IBET	EPDP	ESCP	IEMP	CSIP	EPBI	EEPM	BIPM	IFMS	LT\$	Total
1	Aduthurai	*	*											*					*	*	*			*		5
2	Arundhutinagar	*				*										*	*							*		4
3	Bapatla					*										*	*					*	*			5
4	Brahmavar																									0
5	Chatha	*				*		*			*					*	*		*					*	*	8
6	Chinsurah	*				*	*	*		*		*				*	*						*	*	*	10
7	Chiplima	*		*	*			*	*	*			*			*	*	*	*						*	11
8	Coimbatore	*	*				*	*	*	*	*	*		*		*		*	*		*			*	*	14
9	Cuttack	*	*	*		Gerua	*	*				*	*	*		*							Gerua	*		11
10	Gangavathi	*	*			*		*	*	*				*		*	*		*			*		*	*	12
11	Ghaghraghat	*					*			*		*					*									4
12	IIRR		*	*	*	*	*	*	*	*	*	*	*	*		*		*					*			15
13	Iroishemba	*																								0
14	Jagdapur	*		*	*	*		*	*	*			*			*	*		*	*	*		*	*	*	15
15	Jagtial	*	*	*	*								*		*						*				*	7
16	Karaikal	*				*										*	*								*	4
17	Karjat	*				*										*	*	*		*			*	*	*	8
18	Kaul	*															*								*	2
19	Khudwani	*				*		*			*					*	*								*	6
20	Kurumbapet	*														*								*	*	3
21	Ludhiana	*	*			*		*	*	*	*	*		*					*		*		*	*	*	13
22	Malan	*				*	*	*		*	*					*	*		*			*		*	*	11
23	Mandya	*	*					*		*		*		*		*			*		*		*	*	*	10
24	Maruteru	*	*	*				*	*	*	*	*	*	*		*		*	*	*					*	14

Trial Star Sheet - Kharif 2019 contd...

Sl.No.	Location	PSP\$	PHS	GMS	GMSS	LFST	SBST	MRST	NSN-1	NSN-2	NSNH	NHSN	GMBT	PHSS	GMFM	IBET	EPDP	ESCP	IEMP	CSIP	EPBI	EEPFI	BIPFI	IPMS	LT\$	Total
25	Masodha	*				*		*	*							*	*						*	*	*	8
26	Moncompu	*		*	*		*		*	*		*	*		*				*			*	*		*	12
27	Navsari	*				*	*	*		*						*	*		*		*				*	9
28	Nawagam	*	*			*		*	*			*				*	*		*						*	9
29	Nellore	*		*		*		*					*			*								*	*	7
30	New Delhi	*												*		*	*					*				4
31	Pantnagar	*	*				*	*	*	*	*	*		*					*	*				*	*	12
32	Pattambi	*		*	*	*		*				*	*		*	*			*	*	**		*		*	13
33	Pusa	*					*	*	*							*	*							*		6
34	R. Nagar	*	*			*	*	*	*			*		*					*	*		*		*	*	12
35	Ragolu	*		*	*			*	*				*		*	*					*				*	9
36	Raipur	*					*	*	*			*	*			*	*	*	*		**		*	*	*	14
37	Ranchi	*		*	*			*					*			*	*		*				*			8
38	Rewa	*															*		*						*	3
39	Sakoli	*	*	*	*			*	*				*		*	*	*				*			*	*	12
40	Titabar	*						*	*							*	*				*		*	*	*	8
41	Wangbal																								*	1
42	Warangal	*	*	*	*			*	*				*	*	*	*					*	*			*	12
	Locations	39	14	13	10	19	12	27	18	14	8	14	14	12	6	31	23	6	18	7	14	9	13	20	30	351

Rabi 2019-20

Sl.No.	Location	LFST	SBST	MRST	NSN2	IBET	EPDP	EEPM	YLET	IPMS	Total
1	Aduthurai	*	*	*	*	*	*	*	*	*	9
2	Arundhutinagar										
3	Bapatla										
4	Brahmavar										
5	Chatha										
6	Chinsurah		*			*	*		*	*	5
7	Chiplima					*					1
8	Coimbatore					*					1
9	Cuttack		Gerua			Gerua					2
10	Gangavathi					*		*		*	3
11	Ghaghraghat										
12	IIRR		*								1
13	Iroishemba										
14	Jagdapur										
15	Karaikal										
16	Karjat					*				*	2
17	Kaul										
18	Khudwani										
19	Kurumbapet										
20	Ludhiana										
21	Malan										
22	Mandya										
23	Maruteru		*	*		*	*	*		*	6
24	Masodha										
25	Moncompu							*			1
26	Navsari										
27	Nawagam										
28	Nellore										
29	New Delhi										
30	Pantnagar										
31	Pattambi		*			*			*	*	4
32	Pusa										
33	Ragolu					*					1
34	Raipur					*	*				2
35	R.Nagar		*	*						*	3
36	Ranchi										
37	Rewa										
38	Sakoli										
39	Titabar					*					1
40	Wangbal										
41	Warangal										
	Locations	1	7	3	1	12	4	4	3	7	42

§-PSR and LT Data to be collected for the whole year (January to December)

Coordinated Entomology Trials, *kharif* 2019

Name of the study	: Pest Survey Reports (PSR)
Objectives	: To monitor and report incidence, buildup and outbreaks of insect pests of rice in the region catered by the AICRIP center. Quantification of affected area and intensity of pest damage and impact on yield.
Method	: Visit, survey and surveillance and interaction with local farmers.
Periodicity	: Once in a fortnight. At least six times in a crop season
Target area	: Covering the district where centre is located and 2-3 adjoining districts. In case of pest outbreaks, affected area may be specifically visited.
Essential information	: <ol style="list-style-type: none">1. Specific site & date visited – District, Mandal (Taluk), village (Give specific GPS coordinates).2. Area covered – in multiples of 10 ha3. No. of fields specifically examined4. Variety grown5. Major pest(s) noticed6. Severity of damage (slight, moderate, severe)7. Any other production constraints noticed <i>viz.</i>, drought, flood, diseases etc.
Desirable additional information in respect of severely damaged field(s)	: <ol style="list-style-type: none">8. Age of crop in severely damaged field(s) (in DAT/DAS). Select ten sites randomly representing the whole area and record observations on 10 hills at each site.9. Plant protection measures adopted by the farmer prior to the visit with name & dates of insecticide application.10. Information on fertilizer/fungicide/weedicide application, if any.11. Advice given to the farmer and follow up report if feasible

Submission of report

As early as possible by e-mail (gururajkatti@yahoo.com -), not later than 15th and 30th of each month.

Note: 1) Report may also be based on visit of farmers to the centre with samples of affected plants.

2) Submit report even if there is no appreciable pest damage in the region.

3) If required to visit an affected area, expenditure on POL for the purpose may be claimed with prior approval of the Project Director of IIRR- e-mail request may be made for this purpose to seek permission.

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Pest Survey Report

AICRIP Centre:

Site visited/reported:

Date:

GPS Coordinates:

1. Specific site District, Mandal (Taluk), village		
2. Area covered – in multiples of 10 ha		
3. No. of fields specifically examined		
4. Variety grown		
5. Major pest(s) noticed		
6. Severity of damage (slight, moderate, severe) Please mention the average of observation recorded in ten sites for each pest.		
7. Per cent severity of damage (indicate the extent). Per cent Severity is must for reporting outbreak status of the pest.		
8. Any other production constraints noticed viz., drought, flood, diseases etc.		
9. Age of crop in severely damaged field(s) (in DAT/DAS)		
10. Plant protection measures adopted by the farmer prior to the visit with name & dates of insecticide application		
11. Information on fertilizer/ fungicide/ weedicide application, if any.		
12. Advice given to the farmer and follow up report if feasible		

Please send by e-mail to gururajkatti@yahoo.com latest by 15th and 30th of every month

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 Coordinated Entomology Trials, *Kharif 2019*

1. Host Plant Resistance Studies

Name of the trial	:	Planthopper Screening (PHS) trial
Objectives	:	I. To study the reaction of cultures against brown plant-hopper and whitebacked planthopper with a view to identify the promising material (PHS).
Entries	:	List to be enclosed along with seed material.
A) Field Screening		
Replications	:	One.
Planting date	:	Sowing and planting should be done so as to obtain high planthopper infestation.
Spacing	:	10 x 10 cm.
Age of seedlings at planting	:	3 - 3 1/2 weeks.
Seedlings/hill	:	One.
Check variety	:	Taichung Native 1 (Susceptible).
Plot size	:	Two rows of 10 hills each. Nine rows of test variety alternating with one row of susceptible check TN 1. All around test entries, plant 4-5 infestor rows of tall, susceptible, long duration variety like Mahsuri or Jaya or a local susceptible check
Fertilizer	:	Apply fertilizers according to local recommendations to get higher yields (more N may be top-dressed to get higher infestation).
Chemical control	:	1. Nursery should be protected with suitable insecticide spray at 0.5 kg a.i./ha if other pests are in considerable number.
		2. No control measures should be adopted after transplanting.

Observations:

1. Observe and report planthopper population on 10 hills/entry at 10 days interval from 60 days onwards till 10 days before harvest. Report number of BPH and WBPH/hill separately.
2. Report number of dead and surviving plants per variety first at the time of hopper burn in any of the test varieties followed by another observation prior to harvest.
3. If hopper burn is not observed despite high PH population, record percent tiller mortality in 5 random hills per entry.
4. Report overall damage on 0-9 scale for each entry as described below.

0	No damage.
1	Slight yellowing of a few plants
3	Leaves partially yellow but with no hopperburn.
5	Leaves with pronounced yellowing and some stunting or wilting and 10 -25% of plants with hopper burn, remaining plants severely stunted.
7	More than half of the plants wilting or with hopper burn, remaining plants severely stunted.
9	All plants dead.

(N.B: If plant mortality is due to combined populations of BPH and WBPH and/or other causes, specify them clearly).

Special Instructions: It is important to ensure field reaction through following steps.

1. Erect a polythene sheet barrier of 2.5 feet height all around the planting area within 15 days after planting. For better results it is desirable to plant test entries in longitudinal strips not wider than 2 meters and each strip separately covered around with polythene sheet.
2. Collect adults and nymphs of planthoppers from adjacent areas or green house culture and release them uniformly in polythene confined area on 30, 40, 50 and 60 DAT.
3. Spray 0.002 per cent deltamethrin on infestor/feeder rows 35, 45, 55 and 65 DAT to ensure further build up of the pest population.
4. Population structure as ratio of BPH to WBPH may be furnished when mixed populations prevail in the field
5. **Seed should be collected separately from each culture (5 low damaged hills/culture) which shows very low damage. This seed should be sent to the Principal Scientist & Head, Department of Entomology, IIRR, Hyderabad along with an email intimation to gururajkatti@yahoo.com.**

Wherever facilities are available, the entries are to be tested under greenhouse conditions by adopting standardized technique of mass screening (three replications).

The procedure for mass screening is as follows:

Mass screening:

- ❖ This method involves growing of the test cultures in screening trays/seed boxes of size (50 X 40 X 7 cm).
- ❖ Fill the Seed boxes with well puddled and manure enriched soil and level. Draw 13 equidistant lines horizontally in the box.
- ❖ Draw two vertical lines in the centre of the box cutting the five lines on either side of the middle horizontal line without touching the two boarder lines and middle horizontal lines.

- ❖ Soak the seed of test entries in the petridishes along with susceptible and resistant checks. Keep the soaked seed in a plastic tray and cover with another tray. Next day, remove the water from the petridishes and allow entries to sprout.
- ❖ Sow 20 test entries in the test entry lines by using forceps. Sow two border rows with susceptible check, TN1 and middle row with resistant check, PTB 33 for BPH and MO1 for WBPH. Sow at least 20 seeds of test entries per each line and 40 seeds of susceptible and resistant checks per line. This layout minimizes the chances of escape of the test entries from insect attack.
- ❖ Keep these seed boxes in big aluminium or fibre trays in the plant growth chambers. 10 days (WBPH) -12 days (BPH) after sowing when the plants are of 3-leaf stage, transfer these seed boxes to the screening chambers and cover with cages made of mylar sheet.
- ❖ Release required number of first instar nymphs on the seedlings so that each seedling gets 6-8 nymphs. Cover these mylar cages with plastic mesh so that the insects cannot escape. This infestation is sufficient to kill the susceptible check in 6-7 days. Monitor plant damage regularly.

When TN1 plants on one side show severe damage, rotate the tray by 180° for even reaction. When 90% of plants in the susceptible check, TN1 on both sides are killed, the damage rating of the entries is to be done. Score all the plants in a test entry and checks and score individually, total and average. Score the entries according to Standard Evaluation Scale (SES 2014) on 0-9 scale developed by IRRI

Reference: IRRI (International Rice Research Institute). 2014. Standard Evaluation System for rice (SES), 5th edition. Los Baños (Philippines): International Rice Research Institute

❖).

0	None of the leaves yellow or dried
1	One bottom leaf yellow/dried
3	One or two leaves yellow or one leaf dried
5	One or two leaves dried or one leaf healthy
7	All leaves dried/ yellow but stem green
9	Plant dead

Note:

- ❖ If, as in the past years, PH incidence at your location is consistently high during *rabi* than *kharif*, the trial may be conducted during *rabi*
- ❖ If hopper burn evaluated on visual basis– Kindly indicate the same

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Coordinated Entomology Trials, Kharif 2019

Name of the trial	: Gall Midge Screening (GMS) / Gall Midge Special Screening (GMSS).
Objectives	: To assess the reaction of advanced cultures/donors against gall midge.
Entries	: As per list to be enclosed along with the seed material.
Replications	: One
Plot size	: 1 row of 20 hills per variety/culture.
Planting date	: One late planting (4 weeks later than normal planting). The idea is to adjust the time of planting in such a way so as to synchronize the most vulnerable stage of the plant with peak emergence of the insect.
Spacing	: 15 x 15 cm.
Age of seedlings	: 3 - 3 1/2 weeks
Seedlings/hill	: One
Fertilizer	: Apply fertilizers according to local recommended practice for obtaining high yields (more N may be top-dressed to get higher infestation).

Observations :

- 1) At 30 and 50 DAT, observe all plants to report total plants (TP) and gall midge damaged plants (DP).
- 2) Also record from a maximum of 10 damaged plants/entry the number of total tillers (TT) and silver shoots (SS).

Special Instructions:

1. **Seed should be collected separately from each culture (5 damage free hills/culture) which show nil or very low incidence of gall midge. This seed should be sent to the Principal Scientist & Head, Department of Entomology, Indian Institute of Rice Research, Rajendranagar, Hyderabad - 500 030, Telangana, along with an email intimation to gururajkatti@yahoo.com.**
2. No insecticide should be applied in this trial.
3. No weedicide should be applied in this trial.
4. In case, pest population build-up is seen during post-tillering stage, induce fresh tillering in 50% of hills of each entry by cutting the tillers at water level and record the damage at peak periods.

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Name of the trial	:	Leaf Folder Screening Trial (LFST)
Objective	:	To evaluate entries / breeding lines against leaf folder to identify the promising material.
Entries	:	As per the list enclosed along with seed material
Plot size	:	1 row of 20 hills per entry
Planting dates	:	Sowing and planting dates should be adjusted so as to coincide with high leaf folder infestation
Spacing	:	20 x 15 cm
Age of seedling	:	3 – 31/2 weeks
Seedlings per hill	:	Two
Check varieties	:	Taichung Native 1 (Susceptible check) & W 1263 (resistant check)
Fertilizers	:	Apply fertilizers according to local recommendations to get higher yields. Also apply additional 40kg Urea/ha on 30, 40 & 50 DAT to get higher leaf folder infestation.
Methodology	:	At 25 DAT, cover these entries with nylon net and release leaf folder adults. Collect adults from neighbouring fields or laboratory/glass house culture. Release adults two times, once at 40 DAT and second at 60 DAT @ 100 adults per release. In locations where the leaf folder adult population occurrence is delayed due to climatic variations or other factors, adults may be collected as and when available but preferably release before booting stage. If it gets delayed, releases may be discontinued. Dip cotton in 20% honey solution and place it with a pin inside the net as adult food. Let the adults remain inside the net to lay eggs for a week and then remove the net.
Observations	:	Take observations twice, at 60 DAT and 80 DAT preferably. In case of delayed releases, observations are to be taken 20 days after release. In each entry, select 10 plants at random. Count the total number of leaves and damaged leaves (consider as damaged leaf only if one-third of the leaf area is damaged). Calculate per cent damaged leaves in each entry.
Special Instructions	:	Do not apply insecticides in the main field.

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- Name of the trial : **Stem Borer Screening Trial (SBST)**
- Objective : To evaluate entries / breeding lines against stem borer to identify the promising material.
- Entries : As per the list enclosed along with seed material
- Replications : One
- Plot size : 2 rows of 20 hills per entry with one skip row between entries
- Planting dates : Two planting dates
One normal planting and the second one 15 days after the normal planting
(Accordingly the two sowing dates may be fixed to coincide with peak stem borer incidence of your area)
- Spacing : 20 x 15 cm
- Age of seedling : 3 – 31/2 weeks
- Seedlings per hill : One
- Check variety : PB1, TKM 6, W 1263 and Sasyasree
- Fertilizers : Apply fertilizers according to local recommendations to get higher yields (more N may be top dressed to get higher infestation).
- Methodology : Stem borer infestation may be augmented by pinning of the yellow stem borer egg mass (at black head stage) collected from greenhouse at maximum tillering phase and at booting stage of crop growth.
- Observations :
 - Immediately after transplanting if there's any stem borer incidence count the number of hills that are affected and also for the recovery of the plants.
 - Count the total number of tillers and number of dead hearts (DH) on least 10 hills/entry at 30 DAT or 50 DAT.
 - Also, record total panicle bearing tillers and white ears separately from 10hills/entry at early flowering stage and **prior to harvest.**
 - Grain yield from 5 infested hills. to be taken separately.
 - Stubbles – Count the no. of surviving larvae in three individual infested hills, separately.
- Special Instructions : Do not apply insecticides in the main field.

Damage in the check varieties is important for the trial to be considered as a valid test.

Zero white ear damage in an entry to be confirmed under sufficient pest pressure and are not escapes.

N.B: Record data separately for each of the stages

Send the seeds from 10 best entries as per your evaluation to the Principal Scientist & Head, Department of Entomology, Indian Institute of Rice Research, Rajendranagar, Hyderabad - 500 030, along with an email intimation to gururajkatti@yahoo.com.

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Name of the trial	: Multiple Resistance Screening Trial (MRST)
Objective	: To note the reaction of promising advanced cultures against insect pests with a view to identify multiple resistant cultures.
Entries	: As per list to be enclosed along with seed material.
Replications	: Unreplicated
Planting dates	: Two Staggered sowings and plantings. Planting may be done to coincide with peak pest incidence of your area
Spacing	: 20 x 15 cm
Age of seedlings	: 3 - 3 1/2 weeks.
Seedlings/hill	: One
Check variety	: Taichung Native 1 (Susceptible).
Plot size	: One row of 20 hills each with one skip row between cultures.
Plot arrangements	: Single row of check variety should be included after every 10 varieties/cultures.
Fertilizer	: Apply fertilizers according to local recommendations to get higher yields (more N may be top dressed to get higher infestation).

Observations :

- Record observations on any two major pests only.
- Minor pests when above ETL at any stage of crop growth may also be recorded
- Whorl maggot/leaf folder/hispa/blue beetle : Count the total number of leaves and number of damaged leaves on at least 10 hills/variety or culture at random at 30 and 45 DAT and at peak infestation.
- Gall midge: Count total number of plants and number of damaged plants (bearing silver shoots) on 30 DAT and 50 DAT. Report percent plant damage and percent silver shoots.
-

- Stem borer: Count the total number of tillers and number of dead hearts on at least 10 hills/ culture at 30 DAT or 50 DAT. Also, record total panicle bearing tillers and white ear heads from 10 hills/variety **prior to harvest.**
- Planthoppers and leafhoppers: Report average insect population/hill based on 10 hills/entry along with hopper burn (when observed) and overall plant damage on 0-9 scale as detailed in PHS trial. Greenhouse evaluations wherever feasible are to be done.
- Thrips: Record the damage on 0-9 scale at seedling and tillering stages of crop growth as detailed below:

0	No damage
1	Rolling of terminal 1/3 area of 1 st leaf.
3	Rolling of terminal 1/3 - 1/2 area of 1 st and 2 nd leaves.
5	Rolling of terminal 1/2 area of 1 st , 2 nd and 3 rd leaves, yellowing of leaf tips.
7	Rolling of entire length of all leaves, pronounced yellowing.
9	Complete plant wilting, followed by severe yellowing and scorching.

- Any other pests: Record either pest population/plant or percent damage if pest has caused significant damage. Specify the pest.

Special Instructions:

- Do not apply any insecticide either in nursery or in the main field.
- **Efforts may be made to build up the pest populations for better identification of the resistant/tolerant entries.**
- Specify damage causing pest for each column or observations **along with the age of the crop.**
- Stem borer infestation may be augmented by pinning of the yellow stem borer egg mass (at black head stage) collected from greenhouse at booting stage of crop growth. Similarly, augment other pest populations as indicated in respective pest screening trials.
- Report data only against those pests where pressure was moderate or high.
- The damage units for each pest damage may be clearly specified

N.B: Record data separately for each of the pests.

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Name of the trial	:	National Screening Nurseries (NSN)
Objective	:	To note the reaction of advanced/initial yield trial entries against insect pests.
Entries	:	There will be four sets of NSN. NSN-1 , consists of AVT (Advanced Variety Trials) entries. NSN-2 , consists of IVT (Initial Variety Trials) entries. NSN-(Hills) consists of AVT-hills entries NHSN (Hybrids) consists of IHRT entries
Replications	:	One.
Planting date	:	Adjust time of planting so as to catch up with peak pest pressure.
Spacing	:	20 x 15 cm.
Age of seedlings	:	3 - 3 1/2 weeks.
Seedlings/hill	:	One.
Check variety	:	TN 1
Plot size	:	Each entry one row of 20 hills.
Fertilizer	:	Apply fertilizers according to local recommendations to get higher yields (more N may be top dressed to get higher infestation).

Observations :

- 1) Record observations on two major pests only.
- 2) Refer instruction sheets of earlier trials *viz.*, PHS, GMS, LFST, SBST and MRST for detailed guidelines to record pest incidence/damage.
- 3) Entries may be scored on 0-9 scale as per Standard Evaluation System of IRRI, Philippines. **If SES is not followed, please indicate that it's done by visual scoring on a relative basis.**

N.B: Record data separately for each of the pests and indicate clearly units of observation, pest involved and time of recording data.

Special Instructions:

- Do not apply any insecticide either during nursery or in the main field.
- Evaluations may be carried out under greenhouse conditions at the identified centres for the specified pest.

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2. Insect Biotype Studies

Name of the trial	:	Gall Midge Biotype Trial (GMBT)
Objectives	:	To monitor prevalence, distribution and occurrence of gall midge biotypes within the country.
Differentials	:	As per list to be enclosed along with the seeds.
No. of plantings	:	Late planting to catch up the maximum infestation.
Plot size	:	1 row of 20 hills per variety.
Spacing	:	15 x 15 cm.
Age of seedlings	:	3 - 3 1/2 weeks
Seedlings/hill	:	One
Fertilizer	:	Apply fertilizers according to local recommended practice for obtaining high yields (more N may be top-dressed to get higher infestation).

Observations : 1) At 30 and 50 DAT, examine all plants to report total number of plants and gall midge damaged plants.

2) Also record from a maximum of 10 damaged plants the number of total tillers and silver shoots.

Special Instructions:

Seed should be collected separately from each culture which showed nil incidence of gall midge. **Seed should be sent to the Principal Scientist & Head, Department of Entomology, Indian Institute of Rice Research, Rajendranagar, Hyderabad - 500 030, Telangana. along with an email intimation to gururajkatti@yahoo.com.**

- No insecticide should be applied in this trial.
- No weedicide should be applied in this trial.
- In case pest population build-up is seen during post-tillering stage, induce fresh tillering in 50% of hills of each entry by cutting the tillers at water level and record the damage at peak damage.

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Name of the trial : **Gall Midge Population Monitoring Trial (GMPPM)**

Objectives : To monitor the virulence pattern of gall midge population against select donors.

Differentials :
1. Purple variety (S. Check)
2. RP 2068-18- 3-5 with *gm3* gene for resistance
3. Aganni with *Gm8* gene for resistance
4. W1263 with *Gm1* gene for resistance

Experimental Procedure :

1. Raise nurseries of the differentials (in plastic/GI trays of suitable size) 2 weeks prior to anticipated peak population of gall midge at your location.
2. When seedlings are 1 week old transplant them to about 250 small plastic/clay pots of about 10 cm diameter and 8-10 cm height holding 500 gm soil. Each pot should have 4 hills and each hill with 5 seedlings. Each hill in a pot represents one variety. Label each hill in all the 250 pots. You need 1000 labels. Plant each variety – at predetermined equidistance spots in clockwise order of Purple, RP 2068-18-3-5, Aganni and W1263 (Fig. 1)
3. Take precautions to protect the plants from natural infestation by gall midge by keeping the pots in a net house or in well lighted cages. Avoid exposing plants to electric light source during night times.
4. On the day of infestation, cover each pot with a clear plastic bag (available in any general store). Each bag should just fit the pot at the upper rim. You may use a rubber band or thread to tie, if necessary. Height of bag should be at least 15-20 cm to leave enough space above the plants.
5. Plants should be at least 2 week old and/or of three leaf stage on the day of infestation.
6. To infest each pot, collect one female insect at a light point located near any GM infested plot on the farm. Insects can be collected more easily during peak infestation period between 7.00 and 9.00 pm. Release the insect on to the pot in the bag through a small slit. Care must be taken **to infest each pot with one female only** and seal the slit to prevent escape of the insect.

7. To facilitate infestation of all 250 pots on one day, transport the pots covered with transparent plastic bags to the collection site in the evening itself. Use an appropriate aspirator to collect insect by gently sucking into the tube and then release it through the slit into the bag by gently blowing out.
8. Keep the infested pots covered with plastic bag back in the net house/cage for two days. On third day, remove the bags, water the plants and provide extra humidity for two more days for egg hatching and maggot establishment. This can be done by a humidifier or by periodic (every 30 mins.) spraying of water using a clean plastic atomizer. Alternatively, keep the pots covered with new plastic bag for one more day after watering the pots.
9. Plants are taken care for 3 more weeks until galls develop.

Data recording:

1. When differentials in all the pots show galls, record for each pot, number of gall midge damaged plants for each of the differentials. Record number of galls in Purple variety, RP 2068-18-3-5, Aganni, and W1263.
2. Record sex of the insect emerging from galls for each pot. This can be best done by again covering the pots having silver shoots with the polythene sheet prior to adult emergence and noting the sex of emerging insect. Alternative is to examine the puparium left in the gall exit hole under binocular microscope. Female puparium is slightly larger than the male puparium Fig. 2. Generally, if each pot is infested by a single female, all the emerging insects from a pot will be of one sex. Hence, noting the sex for the first few emerging insects will be good enough.
3. Report data in the following format :

Pot No.	No. of galls in				Sex of emerging adults
	Purple variety	RP 2068-18-3-5	Aganni	W1263	
1					
2					
.					
.					
250					

Seed supply: 100 gm of seeds of each differential is being supplied to the concerned centres viz., Jagtial Moncompu, Ragolu, Sakoli, Pattambi and Warangal.

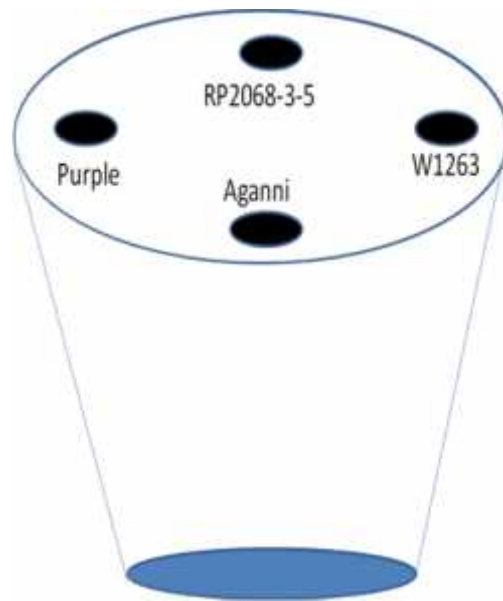


Fig:1 Picture depicting planting of differentials for evaluation in GMPM trial

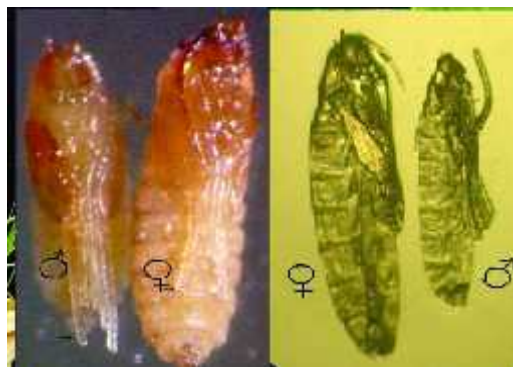


Fig2: Puparia of gall midge

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Name of the trial	:	Planthopper Special Screening (PHSS) Trial
Objectives	:	To monitor virulence in BPH populations (PHSS)
Entries	:	List to be enclosed along with seed material.

Greenhouse Screening

The procedure for mass screening is as follows:

Mass screening:

- ❖ This method involves growing of the test cultures in screening trays/seed boxes of size (50 X 40 X 7 cm).
- ❖ Fill the Seed boxes with well puddled and manure enriched soil and level. Draw 13 equidistant lines horizontally in the box.
- ❖ Draw two vertical lines in the centre of the box cutting the five lines on either side of the middle horizontal line without touching the two boarder lines and middle horizontal lines.
- ❖ Soak the seed of test entries in the petridishes along with susceptible and resistant checks. Keep the soaked seed in a plastic tray and cover with another tray. Next day, remove the water from the petridishes and allow entries to sprout.
- ❖ Sow 20 test entries in the test entry lines by using forceps. Sow two border rows with susceptible check, TN1 and middle row with resistant check, PTB 33 for BPH and MO1 for WBPH. Sow at least 20 seeds of test entries per each line and 40 seeds of susceptible and resistant checks per line. This layout minimizes the chances of escape of the test entries from insect attack.
- ❖ Keep these seed boxes in big aluminium or fibre trays in the plant growth chambers. 10 days (WBPH) -12 days (BPH) after sowing when the plants are of 3-leaf stage, transfer these seed boxes to the screening chambers and cover with cages made of mylar sheet.
- ❖ Release required number of first instar nymphs on the seedlings so that each seedling gets 6-8 nymphs. Cover these mylar cages with plastic mesh so that the insects cannot escape. This infestation is sufficient to kill the susceptible check in 6-7 days. Monitor plant damage regularly.

When TN1 plants on one side show severe damage, rotate the tray by 180° for even reaction. When 90% of plants in the susceptible check, TN1 on both sides are killed, the damage rating of the entries is to be done. Score all the plants in a test entry and checks and score individually, total and average. Score the entries according to Standard Evaluation Scale (SES 2014) on 0-9 scale developed by IRRI

Reference: IRRI (International Rice Research Institute). 2014. Standard Evaluation System for rice (SES), 5th edition. Los Baños (Philippines): International Rice Research Institute

0	None of the leaves yellow or dried
1	One bottom leaf yellow/dried
3	One or two leaves yellow or one leaf dried
5	One or two leaves dried or one leaf healthy
7	All leaves dried/ yellow but stem green
9	Plant dead

Additional studies for PHSS trial:

- ❖ Honeydew test with 30 day old plants – 5 replications
- ❖ Nymphal survival on 30 day old plants – 5 replications
- ❖ Days to wilt on 30 day old plants – 5 replications

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Name of the trial	: Insecticides-Botanicals Evaluation Trial IBET)
Objectives	: To evaluate insecticide and botanical spray modules for efficacy against major insect pests.
Variety	: Any susceptible high yielding variety.
Layout	: Randomized Block Design.
Treatments	: Five
Replications	: Four
Plot size	: 20 – 25 Sq.m
Spacing	: 20 x 15 cm.
Seedlings/hill	: Two.
Age of seedlings at planting	: 3 1/2 – 4 weeks
Time of planting	: Adjust planting time so as to catch peak incidence of insect pests for exposure to treatment application.
Fertilizer	: As per the recommendations for specific area to obtain maximum yields.
Botanicals	: As per the list given in table.
Botanical applications	: At all locations one blanket application of botanical/insecticide should be made at 20-25 DAT. Subsequent applications of botanicals should be given at 10-15 days interval.

Observations :

1. Survey insect populations in experimental plots as well as at light trap at 10 days intervals to judge the time of insecticide application.
2. Silver shoot/dead heart counts on 20 plants based on stratified random sampling should be recorded at 15 days after each application along with total tillers. Follow the same method for white ears at the time of harvest along with total productive tillers.
3. a) Record populations one day before and 3 days after each application in case of external feeders like leafhoppers, planthoppers and hispa on ten random plants.
b) In each plot select 10 random plants and record damaged leaves and total leaves one day before and 7 days after each application.
i) Leaf folder ii) Whorl maggot iii) Rice hispa iv) Other insect pests.
c) Record insect population on 10 hills if ear head bug (gundhi bug) appears in considerable numbers. Also record percent damaged grains.
4. Phytotoxicity if any may be recorded and intimated.
5. Data on natural enemies in 10 hills may be recorded and reported in appropriate format.

Yield data:

Grain yields should be collected from each plot. Exclude 2 border rows on all sides. Mention net plot size and report the yields as Kg/plot.

Special Instructions:

1. Nursery should be protected from insect pests by applying suitable insecticidal spray at 0.5 kg a.i. /ha as and when needed.

2. Individual plots should be separated by bunds and channels to regulate water flow and prevent water movement from one plot to other. Maintain not more than 5-7 cm of water in experimental plots.
3. Efforts should be made to prevent drift between treatments while spraying.
4. Spraying should be done to provide full coverage invariably in the afternoon (after 2pm)
5. In case of direct seeding the experimental plot may be laid out according to the design before seeding. Ensure uniform plant population in all plots.
6. Essential oils should be thoroughly emulsified with any agricultural adjuvant before spray

Note : Details of treatments in Insecticides-Botanicals Evaluation Trial (BIET), *Kharif 2019* and *Rabi 2019-20* are given below.

S.No	Treatment	Details of the treatment	Dose: ml or g/l
1	Botanicals-insecticides	Neemazal 1% EC 25-30 DAT	2.0
		Eucalyptus oil 45-50 DAT	2.0
		Cartap hydrochloride 50% SC 60-65 DAT	2.0
2	Botanicals-Insecticides	Neemazal 1% EC 25-30 DAT	2.0
		Neemoil 45-50DAT	10.0
		Triflumezopyrim 10% SC 60-60DAT	0.48
3	All botanicals	Neemazal 1% EC 25-30 DAT	2.0
		Eucalyptus oil 45-50 DAT	2.0
		Neem oil 60-65 DAT	10.0
4	All insecticides	Chlorantraniliprole 0.4G 25-30 DAT	1.0g/sq.m.
		Cartap hydrochloride 50% SC 50-55 DAT	2.0
		Triflumezopyrim 10% SC 65-70 DAT	0.48
5	Untreated control	Only water spray	

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4. Ecological Studies

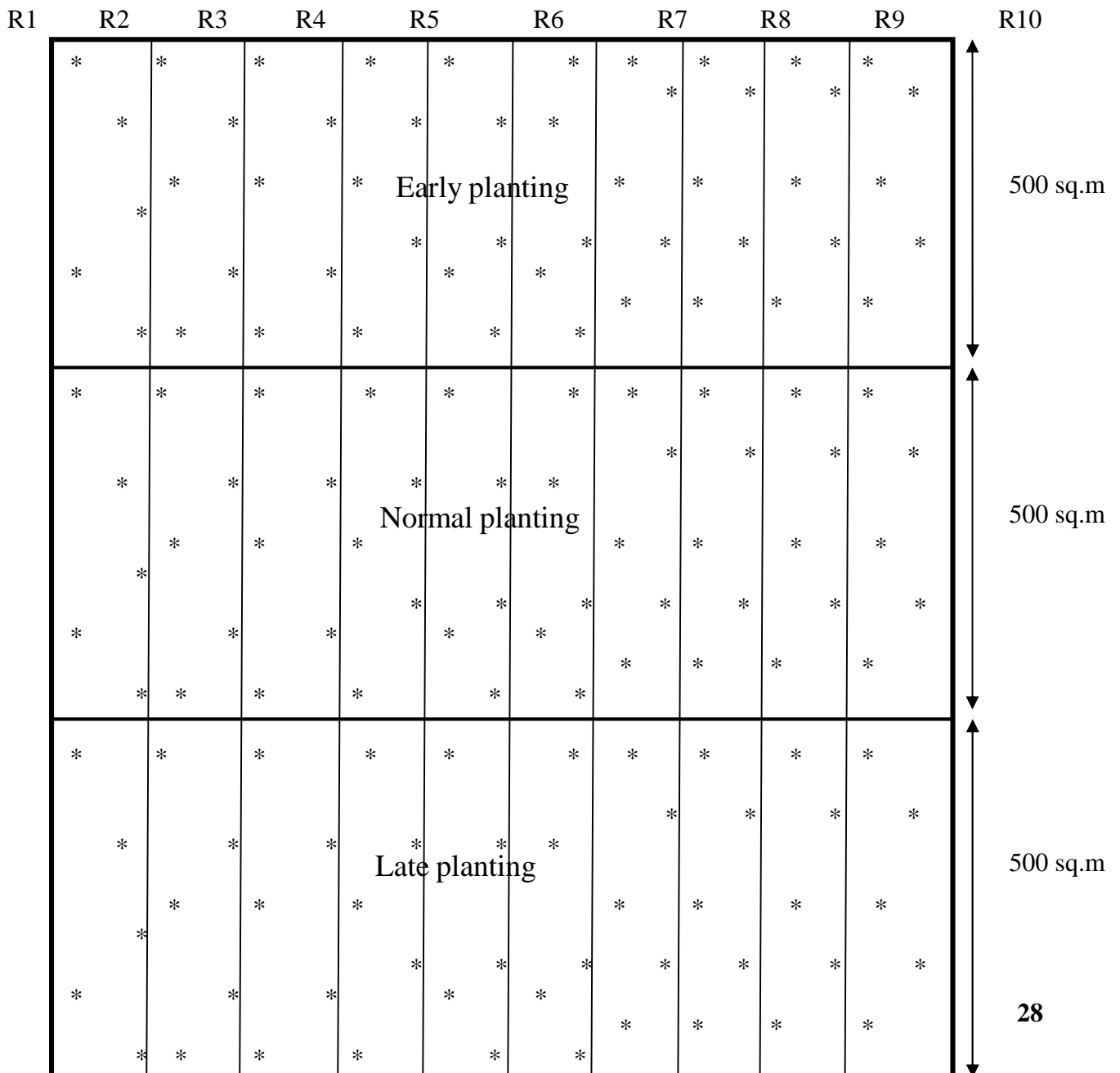
Name of the trial	:	Effect of Planting Dates on Pest Incidence (EPDP)
Objectives	:	To study the influence of date of planting on insect pest incidence and their population dynamics
Variety	:	Any popular high yielding variety
Layout	:	Randomized Block Design
Treatments	:	Three
Replications	:	Ten
Plot size	:	1500 sq.m
Spacing	:	20 x 15 cm.
Seedling/hill	:	Two.
Age of seedlings at planting	:	3½ - 4 weeks
Layout	:	1500 sq.m area is divided into 3 plots of 500 sq.m At each date, planting is to be done in 500 sq.m
Time of planting	:	3 dates of planting 1) Early planting 2) Normal planting 3) Late planting 20 days interval to be kept between the plantings Please note: For each planting, nursery is to be raised separately and transplant same age seedlings on different dates of planting (for example, if 24 day old seedlings were transplanted in early planting, please try to follow the same age i.e., 24 day old seedlings only in normal and late plantings also).
Fertilizer	:	As per the recommendations of specific location
Installation of Data logger	:	Install the data logger at the center of the 500 sq.m field in any date of planting with high pest incidence. Keep it above the water level starting from one week after planting and follow the steps for installation.

Note: Need based application of fungicides and herbicides can be done
No application of insecticides at any stage of the crop

Observations:

- Divide each plot in each planting date (500 sq. m area) into 10 sub-plots of 50 square meter area.
- In each sub-plot, mark **5 hills at random** and record observations on insect pests on **these marked hills** starting from the first appearance of the pest at 10 day interval. At each observation, in **marked hills**, count total number of tillers per hill, number of **dead hearts per hill**, number of **silver shoots**, number of **damaged leaves (specify the pest)**, number of total leaves, total number of panicle bearing tillers and white ears per hill.
- Also record the first appearance of the pest in each date of planting.

LAYOUT FOR EFFECT OF PLANTING DATES ON PEST INCIDENCE (EPDP)



*= hill

Data logger's data recording:

- i) Please take a photograph after installation of the data logger and send it immediately to confirm the method of installation.
- ii) Keep logger interval as three hours so that we get 8 observations each day and after one month of installation, download the data and send the data for verifying the correct recording. Keep the logger till harvest and download the data and send it immediately

Precautions to be taken:

- i) After a heavy shower or wind, please check the data logger and place them straight and in proper position
- ii) Sensor should always be above the water level. If it gets flooded due to heavy rain, please remove immediately, clean with a dry cloth and place them in the field again.
- iii) Don't place them near the irrigation water channels or drainage channels.

Note: *Step wise installation procedure sent separately*

Coordinated Entomology Trials, *Kharif 2019 & Rabi 2019-20*

Name of the trial : Effect of seed coating on insect pest incidence (ESCP)

Collaborative trial with Agronomy (YET 4 - Enhancing the productivity of direct seeded rice with iron coating under different rice ecologies)

Objective : To assess the impact of Iron seed coating on insect pest incidence

Locations (6) : Chiplima, Coimbatore, IIRR, Karjat, Maruteru, Raipur

Treatments, design, plot size, variety and layout are as per the Agronomy technical program. Please consult the Agronomist of your centre. Entomologists will only record observations.

Observations to be taken by Entomologist

In three replications of all the treatments in two sowings (first sowing and fourth sowing), record three observations at 15, 30 and 45 DAS.

Also record pre-harvest/harvest count of panicle bearing tillers and white ears

At each observation, in each replication, select 5 plants at random and record the following:

- 1) Total number of tillers / plant
- 2) Total number of leaves/ plant
- 3) Number of dead hearts/ plant
- 4) Number of galls/ plant
- 5) Number of damaged leaves (indicate the pest- hispa, leaf folder, whorl maggot, thrips, case worm etc./ plant)
- 6) Panicle bearing tillers / plant
- 7) White ears/ plant
- 8) Number of BPH/WBPH/GLH per plant
- 9) Any other pest observed
- 10) Natural enemy count

Coordinated Entomology Trials, Kharif 2019 & Rabi 2019-20

Name of the trial : **Influence of Establishment Methods on Pest incidence (IEMP)**
Collaborative trial with Agronomy (New trial on Long term trial on weed dynamics in mono/double cropped rice systems)

Objective : To assess the influence of different rice establishment methods and weed management practices on insect pest incidence

Treatments : Main plot treatments include 3 establishment methods out of 6 at each location and 4 weed management practices as sub-plots

Locations (8) : Aduthurai, Gangavathi, Ludhiana, Maruteru, Pantnagar, Pattambi, Rajendranagar, IRR

- **Treatments, replications, design, plot size, variety and layout are as per the Agronomy technical program.**
- **Please consult Agronomist of your centre.**
- **Entomologists will record observations in all the plots**

Observations will be taken in 3 establishment methods in all the sub-plots

In three replications of all the establishment methods, record observations at 15 day interval starting from 15 days of planting/sowing.

At each observation, in each replication, select 5 plants at random and record the following:

- 1) Total number of tillers / plant ; 2) Total number of leaves/ plant
- 2) Number of dead hearts/ plant; 4) Number of galls/ plant
- 3) Number of damaged leaves (indicate the pest- hispa, leaf folder, whorl maggot, thrips, case worm etc./ plant)
- 4) Panicle bearing tillers / plant; White ears/ plant
- 5) Number of BPH/WBPH/GLH per plant
- 6) Any other pest observed; Natural enemy count

Coordinated Entomology Trials, *Kharif* 2019 & *Rabi* 2019-20

Name of the trial : **Cropping Systems influence on pest incidence (CSIP)**

Collaborative trial with Agronomy (CA/SM 1 - Conservation Agriculture/ System based management practices in rice and rice based cropping systems (Crop diversification) to utilise resources and enhance the productivity and profitability).

Objectives : i) To study the effect of cropping systems on carryover of insect pests in rice
ii) To assess the effect of *Kharif* rice residue straw mulch on insect pest incidence

Locations (7) : Aduthurai, Maruteru, Karjat, Pantnagar, Rajendranagar, Jagdalpur, Pattambi

- **Treatments, design, plot size, variety and layout are as per the Agronomy technical program.**
- **Entomologists will be taking only observations.**
- **Observations will be taken in all the establishment methods & cropping sequences & residue management plots**

In three replications of all the establishment methods with residue management and cropping sequences, record observations at 10 day interval starting from 15 days of planting/sowing.

At each observation, in each replication, select 5 plants at random and record the following:

- 1) Total number of tillers / plant
- 2) Total number of leaves/ plant
- 3) Number of dead hearts/ plant
- 4) Number of galls/ plant
- 5) Number of damaged leaves (indicate the pest- hispa, leaf folder, whorl maggot, thrips, case worm etc./ plant)
- 6) Panicle bearing tillers / plant
- 7) White ears/ plant
- 8) Number of BPH/WBPH/GLH per plant
- 9) Any other pest observed
- 10) Natural enemy count
- 11) Record yield

Coordinated Entomology Trials, *Kharif 2019 & Rabi 2019-20*

- Name of the trial** : Evaluation of pheromone blends for insect pests of rice (EPBI)
- Objective** : To evaluate various blends and doses of pheromone compounds for monitoring rice leaf folder and pink stem borer
- Replications** : 3
- Plot size** : 1 acre area (these can be placed in any field in the station, seed production plots/ general exhibition plots)
- Treatments** : 4 treatments (traps and lures will be sent along with installation details)
- Locations** : Aduthurai, Coimbatore, Pattambi, Jagdalpur, Navasari, Raipur, Sakoli, Titabar (Leaf folder- 8)
Ludhiana, Pattambi, Warangal, Raipur, Jagtial (Pink stem borer – 5)
- Observations** :1) Observe number of moths caught in each trap at weekly interval
2) Observe the sex of the moths caught in the trap at each observation
3) Also record field damage caused by rice leaf folder and pink stem borer in the field in which traps were installed

Precautions to be taken:

- 1) Always check the trap after heavy rain/ wind and it should be kept erect
- 2) Place it above the crop canopy for pink stem borer (1 ft above the canopy) and below the canopy for rice leaf folder.
- 3) Keep recording the adult catches every week and remove the adults
- 4) Tie the trap with a thread to the bamboo peg for good collection of adults

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5. Biocontrol and Biodiversity Studies

Name of the trial	: Ecological Engineering for Planthopper Management (EPPM)
Objectives	: To use ecological methods to manage planthopper pests
Variety	: Any susceptible high yielding variety
Treatments	1) Ecologically Engineered Plot (EEP) 2) with or without water management (AWD)
Plot Size:	2) Farmers Practice A compact block of 1000 Sq.m. area. for each treatment
Spacing	: 20X20 cm all treatments with alleyway for EEP a must
Seedlings/hill	: Two
Age of seedlings at planting	: 3 1/2 to 4 weeks
Time of planting	: Adjust planting time so as to catch peak incidence of insect pests
Fertilizer	: For FP: As per recommended package of practices specific to each location including insecticides and fungicides as per local farmer practice For EPPM : Addition of vermicompost or any other organic manure Not to exceed 100 Kg N, others as above
Insecticides	: For EEP: Application of granular insecticides in nursery 7 days before transplanting and one granular application at 45 DAT (please note there is minimal application of insecticide in EEP) For FP: Farmers practice which should be recorded.
Methodology for layout	: Divide the entire block into 5 sub-blocks of equal size and demarcate the EEP blocks with alley ways (one skip row) at every 2 m distance. AWD as subplot can be taken in EEP. For FP, sub-blocks are demarked by pegs. Each sub-block will be one replication for observations. There will be a total of 10 blocks
EE interventions	A. Increasing floral diversity <ul style="list-style-type: none">• Increasing floral diversity through locally available flowering weeds/ bund crops that can be a good nectar source. White and Yellow flowers of compositae are reported to attract more wasps. Some plant species recommended are Crops: (<i>Coriandrum sativum</i>), sesamum (<i>Sesamum indicum</i>), greengram, blackgram, cowpea, marigold, Gaillardia, coriander, sunhemp or any bund crop suitable to your region and farmers• Monitor stem borer brood through pheromone traps

YOU CAN PLANT ANY FLOWERING SPECIES SUITABLE TO YOUR REGION

B. Cultural methods

1. Follow line planting with alleyways at every 2 -2.5m
2. Application of vermicompost or any organic manures
3. Mid season draining of water
4. Avoid insecticide spray up to 45 DAT following which 1-2 need based applications may be given

C. Augmenting natural enemies

- Place left over seedlings from nursery, in a small bucket covered with mesh . This prevents hoppers moving out while letting egg parasitoids move into field
- Augment mirids from adjoining area or from glasshouse cultures where available
- Release *Trichogramma japonicum* / *T chilonis* separately or in combination based on presence of stemborer or leaffolder incidence respectively (compulsory in EEP plots)
- Time the release of *Trichogramma* for stemborer based on pheromone catches

Farmers Practice plots : Location specific practices as followed by farmers

Observations

1. Visual counts of hoppers, and any other pest observed on 10 hills/sub-block at 15 days interval
2. Counts of predators such as mirids, spiders coccinellids on these hills simultaneously
3. Drynid parasitisation based on cocoon count.
4. Count other parasitoids such as stem borer egg parasitoids, larval parasitoids such as *Charops* which can be easily seen in the field. Collect stem borer egg masses regularly and observe parasitoid emerging and percent parasitisation
5. Egg baiting for studying egg parasitisation: Potted plant with one or two tillers are exposed to 5 pairs of hoppers for 24 hrs. The adults are then removed and the oviposited plants containing freshly laid planthopper eggs are placed in field for three days near flowering borders. The pots are then brought back to the laboratory for noting egg parasitoids. Dissect the tillers under a microscope for observing parasitized eggs. Unparasitised eggs are creamy white while parasitised eggs are lemon yellow or orange red in colour based on the species of parasitoid. (Fig.)The tillers with eggs should be placed on moist filter paper in petri plates for parasitoid emergence or alternatively, examine one tiller per hill for egg parasite emergence hole and report number per tiller per replication.

6. If hopper damage is seen, score each sub block on 0-9 scale or report per cent plant mortality.
7. Please record cost of cultivation for various practices in the two plots in order to calculate the benefit: cost ratio

- **Please send specimens preserved in alcohol of all parasitoids collected to Dr. Chitra Shanker, Principal Scientist, IIRR, Hyderabad for confirmation and identification.**
- **It is mandatory that you send all egg parasitoids collected from stem borer egg masses pooled into one sample for maintaining a repository at IIRR.**
- **Centres reporting on stem borer species other than YSB and PSB are also required to send specimens of the collected stem borer species.**
- **Refer to:**
http://agropedialabs.iitk.ac.in/i3r/sites/default/files/bc_11sept%20%5BCompatibility%20Mode%5D.pdf for images of various natural enemies



a. White patch showing hopper oviposition



b. Parasitoid emergence holes



c. Healthy eggs



d. Anagrus parasitised eggs



e. Oligosita parasitised eggs

You should compulsorily send

- **a minimum of 20 parasitised egg masses in individual vials.**
- All drynid cocoons and emerging parasitoids
- Coccinellid adults
- Any other larval parasitoids observed

The vials should be forwarded during monitoring visits for species identification and inclusion in the report and to be handed over to Dr. Chitra Shanker, Principal Scientist, IIRR, Hyderabad for identification. The vials should be labeled with location, stage of the crop, season, variety and date of collection.

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Coordinated Entomology Trials, Kharif 2019 and Rabi 2019-20

Name of the Trial **Bio-intensive pest management trial (BIPM)**
Objective

Module	Details of treatments	Practices
BIPM 1	Seed	Seed treatment with <i>Psuedomonas flourescens</i> (@ 10 g / kg seed at the time of sowing or Wet seed treatment @ 10 g / litre of water / kg seed
	Nursery	FYM @ 1 kg / m ² and 100g of rice husk ash / m ² of the nursery bed and mix well with the soil at the time of preparation of the field
	Fertilization	5 tonnes of FYM/ compost/ green leaf manure or 2.5 tonnes of vermicompost as basal + 300-500 kg neem cake / ha half as basal and half as top dressing at active tillering stage
	Pest Management	<ol style="list-style-type: none"> 1. Clipping of rice seedlings before Transplanting 2. Mass trapping of stem borer by installing pheromone traps @ 20 numbers/ ha. 3. <i>Trichogramma japonicum</i> 5 cc egg cards/ha, six times weekly from first week after transplanting 4. <i>T. chilonis</i> for leaf folder management at weekly intervals from 20 days after transplanting 5. Need based application of neem formulations/ biopesticides for other defoliating pests 6. Foliar spray of <i>P. flourescens</i> on the foliage @ 20 g / litre of water. Spraying can be repeated depending on the disease severity.
BIPM 2		Same as above in all respects except application of <i>Trichoderma</i> IIRR strain instead of <i>Psuedomonas</i>
Farmers Practice		General POP with RFD and need based application of insecticides

Observations to be recorded:

- Observations on pest and disease incidence will be recorded on 50 randomly selected hills in each treatment at fortnightly interval.
- Observations on natural enemies like predators and parasitoids by visual or sweep net count and by collection of egg masses for stem borer
- At harvest record yield in each treatment
- Benefit cost ratio will be calculated

PS: Please indicate requirement of *Trichoderma*

Average nutrient composition (%) of major nutrients of some organic manures

Source	N	P ₂ O ₅	K ₂ O
FYM	0.5-0.8	0.4-0.8	0.5-0.9
Compost	0.5-1.5	0.5-1.4	1.4-1.6
Vermicompost	1.00-2.05	0.70-1.90	1.5-2.5
Poultry litter, fresh	1.0-1.8	1.4-1.8	0.8-0.9
Poultry litter, very dry	3.0-4.5	4.0-5.0	2.0-2.5
Groundnut cake	7.3	1.5	1.3
Castor cake	4.3	1.8	1.3
Neem cake	5.2	1.0	1.4
Green manure (on dry weight basis). Moisture % will be 80-85	2.0-2.5	0.4-0.8	0.5-1.0

Note: Based on the average nutrient composition of the organic source used, the Soil Scientists can calculate the quantity of organic manures **based on the N equivalent basis**.

Observations to be recorded:

- Divide each Treatment block into 6 smaller blocks for observation purpose. Observations on pest incidence should be recorded on 10 randomly selected hills in each replication (60 hills/ each treatment) at fortnightly interval.
- At each observation, record total tillers, dead hearts, silver shoots, total leaves, damaged leaves, number of planthoppers/ hill.
- At harvest record yield/m² randomly at 20 points in each treatment.

Observations to be recorded by Soil Scientists:

Soil analysis:

- Initial soil analysis of two blocks separately for all Soil Characteristics like pH, EC, Organic carbon, available NPK status, micronutrient status and important physical properties.
- Final analysis of soils after harvest for all important properties in smaller blocks of each block.

Plant analysis:

- Grain and straw yields at harvest
- Grain analysis for quality parameters (in brown rice and polished rice) along with hulling, milling and head rice recovery.

Note: If quality analysis is not available at the centres, send grain samples to the PI, (Soil Science), IIRR (DRR) immediately after harvesting.

- Grain and straw analysis for nutrient concentration of major nutrients and Zn and Fe

Number of samples: 2 samples in each small block of 6 in two big blocks (2 X 6 X 2=24)

Total number of samples=24 (soil and plant samples).

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Coordinated Entomology Trials, *Kharif 2019 & Rabi 2019-20*

Name of the Trial Objective	Integrated Pest Management (Special) To validate IPM practices from a basket of options available and demonstrate to farmers the management of pests in a holistic way (including insects, diseases and weeds).
Variety	Local popular variety of the region
Plot size	Two blocks of not less than 1 acre for each block.
Replications	5 replications. Divide each block into 5 equal sized units (each unit = one replication)
Treatments	Take 3-5 farmers in each centre/location, each farmer representing a replication with at least 1acre area/farmer as IPM plots. Farmers can be selected from same village or different villages
Details of the treatments	The package of practices to be followed in each block are given below:

Details of treatments	IPM block	FP block
Nursery	<ul style="list-style-type: none"> • Seed treatment with Carbandezim @ 2 g for kg seed. Soak these treated seed overnight in 10 liter water and keep in gunny bag for germination (see attached detailed instructions) • Apply Carbofuran @ 1.1 kg a.i./ ha, 5 days before pulling seedlings from nursery for transplantation. (In gall midge endemic areas) 	As per the local farmers practice. Please record the practices followed by farmers whenever you go for observation/visit.

Main field	<ul style="list-style-type: none"> • Transplant seedlings at a spacing of 20 x 15 cm. • Leave alleyways of 30 cm after every 2 m or 10 rows. • Fertilizers should be applied as per local recommended fertilizer dose. • Apply Butachlor 1.5 kg a.i./ ha within one week after transplanting the crop . • Survey for pest incidence and level of damage at weekly interval starting from 15 DAT. • If BLB symptoms seen between 20 to 30 DAT, split doses of nitrogen may be delayed, particularly the second dose. • At 15 DAT, install pheromone traps with 5 mg lure @ 8 traps/ ha for stem borer monitoring. While installing make sure that the trap remains above the crop canopy. 	<p>As per the local farmers practice</p> <p>Please record the practices followed by farmers whenever you go for observation / visit</p>
30 – 59 DAT	<ul style="list-style-type: none"> • Depending on weed intensity spray post emergence herbicide as given (Pg. No. 5) • Observe bund area and if sheath blight is observed on weeds, go for spray (as given in Pg.No. 6). • N top dressing to be taken up as given in protocol using Leaf Color Chart • Mid season drainage. 	<p>As per the local farmers practice (mention the quantities)</p> <p>Please record the practices followed by farmers when- ever you go for observation/visit</p>
60 – 90 DAT	<ul style="list-style-type: none"> • One spray of Cartap hydrochloride 50 WP @ 600 g / ha at 60 DAT (against stem borer/leaf folder, if incidence crosses ET value). • Need based application of Propiconazole (PI see Pg No. 6). • Mid season drainage should be followed in case of BPH incidence. 	<p>As per the local farmers practice</p> <p>Please record the practices followed by farmers when-ever you go for observation/visit,</p>
> 90 DAT up to harvest	<ul style="list-style-type: none"> • Mark 5 X 5 m² area and take yield, at 5 places (5 repl.) in this block 	<ul style="list-style-type: none"> • Mark 5 X 5 m² and take yield, at 5 places (5 repl.) in this block

	<ul style="list-style-type: none"> • Also record the cost involved for each practice/ operation taken in IPM starting from nursery to harvest to estimate cost of cultivation as given in data sheet 	<ul style="list-style-type: none"> • Also record the cost involved for each practice/ operation taken up by farmers starting from nursery to harvest to estimate cost of cultivation as given in data sheet
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Observations to be recorded:

- Starting from 15 DAT, observations on pest incidence should be recorded on 5 randomly selected hills (each time hills are selected randomly) in each replication (25hills/ each block) at weekly interval. (Total of 25 hills in IPM block & 25 hills in FP block at each observation).
- At each observation, record total tillers, dead hearts, silver shoots, total leaves, damaged leaves, number of planthoppers/ hill as per the data sheet given.

Record disease incidence (% disease severity) against Blast (leaf/neck), bacterial blight and other major diseases.

Record the following weed observations:

- Weed population (number/m²) 30, 60 DAT
- Dry weight (gm/ m²)of weeds at 30, 60 DAT

Grain yield : Record the yield from 5 places of 5 x 5 m area from each replication.

Note: In case of insect/ disease infestation, please follow ETL's and control measures should be taken as per the IPM guidelines/protocol given below. Inform/consult concerned PI/scientist in case of severe infestation or when in doubt about action to be taken.

IIRR IPM team

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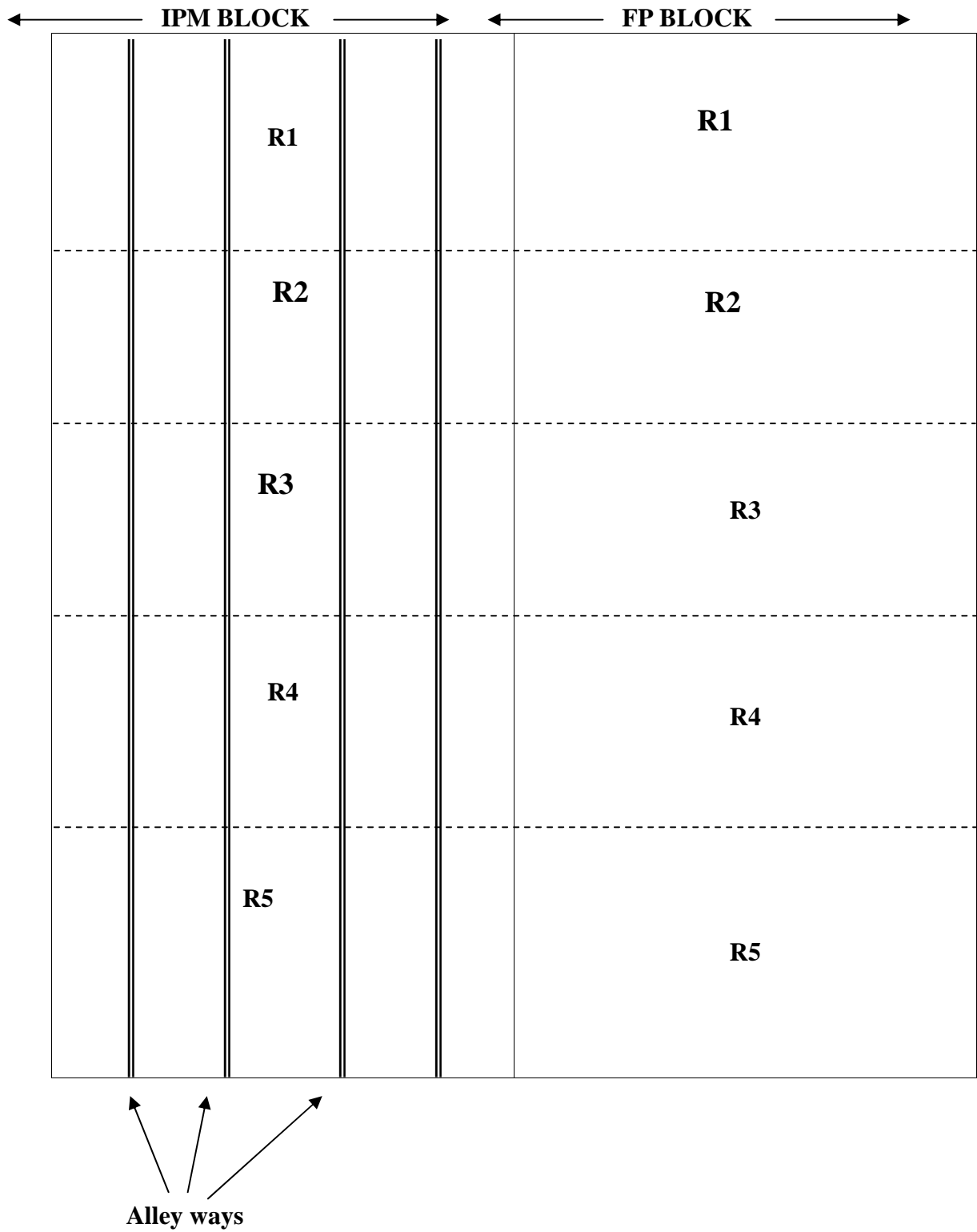
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Note: You can contact anyone at any time

LAYOUT PLAN FOR INTEGRATED PEST MANAGEMENT (SPECIAL) KHARIF 2019



A. Protocol for effective weed management in IPM Special trial (in IPM treatment)

Since the trial is being laid out in irrigated ecology, weed management both in nursery and main field are equally important.

1) Nursery

- i. Maintain water level to avoid weeds
- ii. In weed intense areas, apply Butachlor @25ml/250 m² nursery area or Pretilachlor+safenor @ 60ml/250 m² nursery area application at 8-10 days after sowing seed in nursery beds
- iii. Raising nursery in strips of 1 m wide and leaving water canal of 0.25 m in between will help in intercultural operations

2) Main field: Immediately after transplanting within a week

* Liquid formulation of new herbicides can be applied by mixing with sand or by foliar spray, respectively, within first week after transplanting by following the procedure outlined hereunder.

* Required quantity of herbicide (Butachlor @3 liters/ha or Pretilachlor @1250-1500 ml/ha or Anilophos 1250-1500 ml/ha or Metsulfuron methyl+chlorimuronethyl (Almix)@20g/ha) mixed with fine sand (50kg/ha) and broad casted. Or mixed in 500 liters water/ha and spray by flat Z type nozzle uniformly within 3 to 7 days after transplanting. It is necessary to maintain standing water (2-3 cm water) in the field.

Do not remove water at least 48 hours after application of herbicide.

* **Note** that under thorough land preparation and proper water management conditions this step may not be required. Take a decision on 2nd day after transplanting based on land leveling and water supply status.

Post-emergence application:

*Broad spectrum weed control – Bispyribasodium @ 250ml/ha at 2-3 leaf stage of weeds- spot application or Chlorimuron + Metsulfuron-methyl (Grasses, Sedges and Annual BLW) at 20-25 DAT @ 20 gm/ha

* If Broad leaf weeds predominate, apply 2, 4-D Na salt @ 1250-1500 g/ha at 20-25 DAT

* If grasses predominate, apply Cyhalofbutyl @1000 m/ha at 15-20 DAT or Fenoxaprop p ethyl @ 800-100ml/ ha at 25-30 DAT.

Fertilizer management: Apply top dressing nitrogen based on Leaf Color Chart (modified IIRR - LCC) supplied by IIRR. The instructions to use LCC are given on backside of LCC.

Observation to be recorded under IPM plot as well as in Farmers Practice plots:

Monitor at regular interval weed growth (Group wise no. of weeds i.e., grasses, sedges and broad leaves weeds) in 1 m² area in each replication with the help of a quadrat. Collect all the weeds, dry them in oven at 60^o C for constant dry wet and record dry weight at 20, 40, 60 DAT.

- Weed population (number/m²) 30, 60 DAT
- Dry weight (gm/ m²)of weeds at 30, 60 DAT
- Observe the changes in weed flora

B. Protocol for effective disease management in IPM Special trial (in IPM treatment)

1. Seed Treatment: (can be taken up as prophylactic)

Seed should be treated with carbendazim @ 2 gm/kg seeds (wet seed treatment)

Method: 10 gm of Bavistin should be mixed with 10 litres of water in a bucket and 10 kg cleaned rice seeds should be put in that solution. The chaffs which will float should be removed by hand. Seeds should be mixed with the solution properly with the help of a clean bamboo peg or stick. The seeds should be left in the solution for 24 hours. After 24 hours, the fungicide solution should be decanted and the seeds should be put in a clean and wet cloth bag and should be tied properly. The cloth bag should then be incubated in closed chamber (like cement tank) and should be fully covered with paddy straw. After 24 to 48 hours, the seeds will germinate and the germinated seeds can be used for nursery sowing. Use of hand gloves is must at the time of seed treatment and transfer of seeds from bucket to cloth bags.

Most of the diseases appear in the maximum tillering stage onwards

Blast: If still there is incidence of blast in the nursery, then give one spraying with tricyclazole 75 WP @ 0.6 g/l or iprobenphos 48 EC @ 2g/l or isoprothiolane 40 EC @ 1.5 ml/l or carpropamid 30 SC @ 1 ml/l or carbendazim 50 WP @ 1 g/l or kasugamycin 3 SL @ 2.5 g/l or Epoxyconazole 125 g/l + carbendazim 125 g/l @ 0.5 ml/l.

Sheath blight: Sheath blight in general starts at the tillering to maximum tillering stage. Many cases, it has been noticed that the disease appears near the bund (probably from the infected weed hosts or inoculum present in the infected straw kept in the bunds or the sclerotia floating on water and accumulated near the bunds) and then progresses inwards. Regular surveillance is must and if the initiation of the disease is seen in any parts especially near the bunds, then one spraying can be given especially in the affected area. The sprayings can done with the chemicals like validamycin 3L @2.5 ml/l or propiconazole 25 EC @ 1 ml/l or hexaconazole 5 EC @ 2 ml/l or carbendazim 50 WP @ 1g/l or thifluzamide 24 SC @ 30 g a.i/ ha.

BLB: BLB appears initially in patches and near the shades. If BLB symptoms are noticed, delay the next top dressing.

Brown spot: Under irrigated ecosystem, if the fields are well managed and if fertilizer application is balance, then brown spot will not be a big problem. Moreover, seed treatment with carbendazim will take care of brown spot. However, still if it comes in some of the pockets in plots then, sprayings with chemicals like carbendazim 50 WP @ 1g/l or chlorothalonil 75 WP @ 2g/l or combination of carbendazim (12%) and mancozeb (63%) @ 1.5-2 g/l or mancozeb 75 WP @ 2g/l can be done.

Foot Rot (Bakanae): Generally seed treatment will take care of the seed borne inoculum of the fungus. However, if it is observed then one spraying with carbendazim (0.1%) will take care of the disease.

Stem Rot: Though it is minor disease, it can cause havoc in association with the BPH infestation. If stem rot symptoms are seen, then one spraying with Iprobenphos 48 EC @ 2g/l or carbendazim

50 WP @ 1g/l or thiophanate methyl 70 WP @ 1 g/l or isoprothiolane 40 EC @ 1.5 ml/l can be done.

One need based application (based on the disease history of the location) with 0.1% propiconazole or Nativo (0.4 g/l) around flowering will take care of false smut, grain discolouration and sheath rot diseases.

Economic Thresholds Suggested for application of fungicides

S.No	Disease	ETL
1	Foliar blast	3-5 lesions/leaf
2	Brown spot	2-3 spots/leaf & 2-3 infected plants/ m ²
3	Sheath blight	Lesions of 5-6 mm in length & 2-3-infected plants/m ²
4	Sheath-rot	Lesion length 2-3 mm on sheath & 3-5 infected plants/m ²
5	BLB	2-3 infected leaves/m ²
6	Tungro	1 tungro infected plants/m ² & 2 GLH/hill (in fungus endemic areas)
7	Neck blast	2-5 neck infected plants/m ²

C. Protocol for effective insect pest management in IPM Special trial (in IPM treatment)

Based on the periodic observation compute average pest damage in IPM plot and determine if the damage has crossed Economic threshold level.

Economic Thresholds Suggested for application of insecticides

S.No	Insect pest	ETL
1	Stem borer	10 % dead hearts or one adult moth or one egg mass per sq. m or >30 moths/pheromone trap/week
2	Gall midge	5 percent silver shoots
3	Leaf folder	2 damaged leaves per hill with a live larva.
4	Planthoppers	10 insects/hill at vegetative stage; 20 insects/hill at later stage.

Following information on major pests will help to decide on appropriate IPM interventions

Planthoppers

The pest generally appears 45 to 50 days after planting. Adults and nymphs suck the sap at the base of the tillers, resulting in yellowing and drying of the plants. Infestation spreads in concentric circles and in severe cases the affected field gives a burnt appearance. Provision of alley ways leads to change in micro-climate and helps in monitoring pest population and pesticide application. Regular surveillance is a must starting from 40 DAT. Walk along the alleyways and observe on either side at the base of plants for planthoppers. If the population exceeds ETL, go for suggested measures given. In BPH endemic areas, go for mid season drainage to prevent population buildup.

Stem borers

This pest may appear even in nursery and in main field during any stage of the crop. Adult moths are seen resting on the leaf tip during early hours of the day or egg masses are seen on the stem and leaf. The pest has a patchy distribution resulting in a patch of deadhearts/ whiteears depending on the stage of the crop.

Installing pheromone traps for monitoring the pest is effective way of tracking the pest. Install 8 traps/ha with 5 mg lure such that trap remains above crop canopy. The trap catches are monitored at weekly interval to know the pest buildup. When trap catches exceed 30-35 moths/trap/week, go for suggested measures. Change the lure after 25 days as it loses its effectiveness.

Gall midge

The pest may appear in the nursery or in the main field up to active tillering stage. Galls or silver shoots appear after 4 weeks of adult appearance and egg laying. If pest damage exceeds ETL, resort to control measures as suggested.

Defoliators

Most of the defoliators like leaf folder, case worm, green horned caterpillar, skipper, semi-looper appear immediately after transplanting. Go for regular scouting and only if pest damage exceeds ETL, go for suggested control measures.

D. Operational guidelines for implementing IPM (Special) trial

It is envisaged that IPM (special) trial may be implemented in ‘On-line real-time’ mode. Hence it is essential that all the team leaders of the concerned AICRIP centre’s are in touch with IIRR team and coordination unit on almost daily basis.

IPM is obviously a knowledge intensive technology and its impact depends on timely and informed decisions. Periodic surveillance at weekly interval is the core activity of the Trail and needs to be religiously followed. It is desirable that entire team of scientists visit the experiment site together, as often as possible, during this surveillance. If not, at least a representative of the scientist may accompany the team. .

During any of the surveillance, if the pest damage crosses threshold, IPM interventions need to be decided within 24 h in consultation with the IIRR team. If response is not available from IIRR within this time, local decision may be taken and IIRR be informed of this. Impact of such specific intervention needs to be monitored through subsequent surveillance visit.

It is also important to timely record and report farmer’s practices being followed in FP plots. This information may also be forwarded to IIRR unit.

Coordinated Entomology Trials, *Kharif* 2019 & *Rabi* 2019-20

7. LIGHT TRAP COLLECTION OF INSECTS (LT)

Objective : To monitor on long term basis fluctuations in the populations of insect pests and their natural enemies.

Light Trap Design : Old light trap of the centre to be continued (Mention the type of light trap installed, type of bulb and wattage of bulb used) – Please note,

❖ 200 W incandescent bulb should be used

Reporting data:

- No. of insects collected in **each trap** be recorded **separately daily**, focusing on major insect pests and natural enemies of your region.
- Send raw data for entire year using **MS Excel Data sheet template** for light trap data for processing at DRR
- Light trap data are needed for the **entire year** though there may be a single rice crop at your centre.
- Mention the prevailing cropping system in the area

Additional Information Report the date of planting of rice crop in the adjacent area of the light trap, specify variety and growth stage for each month.



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